The Preparation of 1 β ,11 α -Dihydroxy-steroids by Microbiological Hydroxylation

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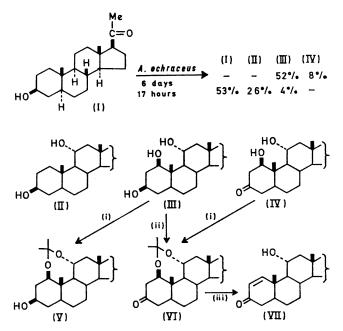
Summary Incubation of 3β -hydroxy- 5α -pregnan-20-one with the fungus Aspergillus ochraceus gives the 18.11adihydroxy-derivative in 52% yield.

THE extensive literature on the fungus Aspergillus ochraceus shows that it generally introduces one hydroxy-group into the steroid nucleus, producing 11a-hydroxy-compounds from a wide range of oxygenated substrates.¹ If access to the 11-position is impeded, or if the size of the 13-alkyl group is increased, hydroxylation occurs at alternative sites.^{1,2} With another fungus, Calonectria decora, there is a predilection for dihydroxylation to give products in which the hydroxy-groups are well separated from each other.³ From these and numerous similar observations it might be inferred that the presence of one oxygen group (whether originally present in the substrate or introduced microbiologically) inhibits entry of a second group into a position which brings the two groups into close proximity. Thus, 11 α - or 1 β -hydroxylation by Absidia orchidis have been regarded as mutually exclusive.3c

We now report that incubation of 3β -hydroxy- 5α -pregnan-20-one (I) with A. ochraceus gives the 1β , 11α -dihydroxyproduct (III) in satisfactory yield. [The conditions³ are reasonably forcing, but not unusual. Each incubation flask contained a culture of the fungus growing vigorously in a corn-steep nutrient (200 ml), to which the steroid (40 mg) was added as a solution in dimethyl sulphoxide. The flasks were swirled at 25° for 6 days.] The product (III) is readily converted into an acetonide (V) whose formulation as a 1,11-isopropylidenedioxy-compound follows from n.m.r. examination:⁴ the remarkable stability of this derivative (which is unchanged by boiling with 2Nhydrochloric acid in dioxan) is paralleled by the very strong 1,11-hydrogen bonding of the parent compound (v_{max} 3320 cm⁻¹). Treatment of the 3-keto-acetonide (VI) with acid afforded the known 11a-hydroxy-diketone (VII)⁵ by the expected β -elimination of the 1-alkoxy-group.

The product (III) is formed by a sequential rather than by a concerted introduction of the two hydroxy-groups. A short hydroxylation period gives mainly the mono-(11a)-hydroxy-derivative (II);⁶ incubation of this compound leads to the dihydroxy-product (III). The results do not fit the kinetic expression for normal consecutive reactions. For example, in parallel experiments using the starting material (I) and the monohydroxy-derivative (II),

the former gives the product (III) more quickly. We incline to the view that enzyme induction⁷ occurs in the present system, and that the rates of the reactions depend upon the speed with which the required enzymes are produced.



Reagents i, H2SO4-Me2CO; ii, H2CrO4-Me2CO; iii, 2N-HCldioxan, reflux.

The course of the hydroxylation of 3β -hydroxy-20-oxo- 5α -pregnanes with A. ochraceus is profoundly influenced by structural variations in the substrate. Under the conditions used here 1β , 11α -dihydroxylation occurs with the 16β -methyl derivative, the 16-17-olefin, and the related 16α , 17α -epoxide, but not with the 17α -alcohol or the 16methyl-16,17-olefin. The present work provides an easy route to 1-dehydro-3-oxo-11a-hydroxy-compounds, and applications to the preparation of physiologically active compounds can be envisaged.

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